EXOGENOUS APPLICATION OF POTASSIUM SILICATE FOR GROWTH ENHANCEMENT AND SALT STRESS ALLEVIATION IN CORIANDER (CORIANDRUM SATIVUM L.)

RITI THAPAR KAPOOR* AND ARTI YADAV

Plant Physiology Laboratory, Amity Institute of Biotechnology, Amity University Uttar Pradesh, Noida-201 313, India

Keywords: Coriandrum sativum, Growth, Potassium silicate, Salt stress

Abstract

Salinity is a major threat to agriculture and its adverse effect on farming land is expanding day by day. In the present study effects of potassium silicate on germination and biochemical parameters of *Coriandrum sativum* under salt stress were studied. The maximum seed germination (94%) of coriander seeds was observed with potassium silicate. Total chlorophyll content showed the following order: PS > C > NaCl (2mM) + PS > NaCl (4 mM) +PS > NaCl (6mM) +PS > NaCl. Potassium silicate enhanced biochemical components such as sugar, proline, protein and total antioxidant contents in coriander seedlings. The maximum total antioxidant content (46%) was observed in NaCl (6mM) + PS treatment. Hence, potassium silicate acts as a growth stimulating agent and can be used as fertilizer for coriander plants under salt stress.

Introduction

Soil salinity is one of the major abiotic stress responsible for decline in crop productivity at global level (Zhu *et al.* 2019). Approximately 40% of cultivable land is affected by salinity (Hafez *et al.* 2021) and saline area is increasing annually as per the rate of 10% due to change in climate conditions, poor cultural practices, utilization of chemical fertilizers and irrigation by using saline water (Abdel Latef *et al.* 2021). Salinity has already affected almost 954 million hectares of the world's total land area (Saddiq *et al.* 2021). Salt stress adversely affects morphological, physiological and biochemical parameters of plants (Farhangi-Abriz and Torabian 2018). Salinity enhances uptake of Na⁺ and Cl⁻ from the soil and reduces transport of essential nutrients and generates osmotic and nutritional components disturb osmotic balance, resulting in physiological drought which prevents water uptake in plants (Riaz *et al.* 2019). Various approaches such as development of salt resistant crops, use of plant growth-promoting bacteria, endophytes have been used to overcome the negative impacts of salt stress for sustainable crop production. All the above mentioned methods are expensive, time consuming and not economically feasible at large scale.

According to Zargar *et al.* (2019), silicon is a multitalented element and can be used as a fertilizer for enhancing crop production. The uptake of potassium and calcium was also improved over sodium absorption with silicon utilization and enhanced the growth of rice, sunflower and sorghum (Hurtado *et al.* 2020). The use of silicon decreased Na⁺ translocation from roots to shoots and leaves of chickpea plant (Garg and Bhandari 2016).

Coriander (*Coriandrum sativum* L.; belonging to Apiaceae), an annual herbaceous plant is most widely used due to its nutritional as well as medicinal properties. Coriander seeds have been used as a spice or traditional medicine for the treatment of indigestion, nausea, dysentery whereas its leaves stimulate appetite (Laribi *et al.* 2015). The presence of essential oil in coriander seeds

^{*}Author for correspondence: <rkapoor@amity.edu>.

shows antimicrobial, anti-inflammatory, antiseptic, anticancerous and free radical scavenging properties (Chahal *et al.* 2017). Application of silicon can be considered as one of the promising methods to improve plant resilience under salt stress (Almeida *et al.* 2017). No reports are available in literature about the impact of potassium silicate for salt stress mitigation in coriander. Therefore, present investigation was carried out to study the role of potassium silicate on growth and biochemical parameters of coriander under salt stress.

Materials and Methods

Coriandrum sativum L. variety Kasturi seeds were procured from seed agency of Ghaziabad, India. Different concentrations of NaCl (2 mM, 4mM and 6 mM) and 2 mM Potassium silicate (K₂SiO₃) were used for the treatment. The coriander seeds were surface sterilized with 0.01% HgCl₂ solution, then washed with distilled water. Thirty seeds were divided into three replicates of 10 seeds and each were immersed in 10 ml of different concentrations of NaCl solution and potassium silicate for five hrs. Petri dishes were kept in a growth chamber under temperature ($25 \pm 2^{\circ}$ C), photoperiod 16/8 hrs and photon flux density was kept 240 µmol m⁻²s⁻¹ for 10 days. Petridishes were kept moist by adding different concentrations of salt, potassium silicate or distilled water as and when required according to the treatment. Different growth characteristics of coriander such as germination percentage, relative germination rate and germination index were determined by the following formula of Li (2008). The radicle and plumule length were measured with a measuring scale.Vigour index of the coriander seedlings was estimated by the method of Abdul - Baki and Anderson (1973). The fresh weight of coriander seedlings was measured ten days after seed sowing. After that, seedlings were oven dried at 65^oC for 72 hrs and dry weight was estimated. RWC was calculated by the modified method of Barrs and Weatherly (1962).

The estimation of chlorophyll a, b and total chlorophyll was done by the modified method of Misyura *et al.* (2013) and carotenoid content was analyzed by Kirk and Allen (1965). Total sugar content was analyzed by the method of Hedge and Hofreiter (1962). Proline content was assessed by the method of Bates *et al.* (1973). The protein content was measured by the method of Lowry *et al.* (1951) by using BSA standard curve. The total antioxidant content in coriander seedlings was evaluated by Prieto *et al.* (1999). Treatments were organized with three replicates in randomized block design. The data were determined by using ANOVA and SPSS software. Mean of treatment was assessed by DMRT at p < 0.05.

Results and Discussion

Potassium silicate treatment showed maximum seed germination (94%) in coriander seeds whereas (89%) was reported in control. Relative germination rate and germination index were more with potassium silicate as compared to other treatments. Maximum (84%) reduction in seed germination was observed with salt treatment (Table 1). Similarly, silicon significantly increased the rate of seed germination in tomato (Shi *et al.* 2014).

Relative water content and vigour index reflected the following trend: PS > C > NaCl (2 mM) + PS > NaCl (4mM) + PS > NaCl (6 mM) + PS > NaCl (Table 2). The present results reflected the same trend with the work of Wu*et al.*(2015) who found the same enhancement in the root and shoot growth in cucumber and tomato by applying silicon under salt stress. Liu*et al.*(2015) reported that addition of silicon improved water uptake in sorghum by up-regulation of aquaporin genes in roots. Silicon improved root growth by promoting Casparian band formation and biosynthesis of suberin and lignin which may check Na⁺ ions entry into the transpiration stream (Guerriero*et al.*2016).

Treatment	Germination (%)	Relative germination rate(RGR)	Germination index(GI)
Control	89 ± 0.62^{a}	-	8.9 ± 0.21^{a}
NaCl (6 mM)	14 ± 0.05^{d}	$0.16\pm0.02^{\rm c}$	1.4 ± 0.10^{d}
PS (2mM)	94 ± 0.84^{a}	$1.06\pm0.59^{\rm a}$	9.4 ± 0.32^{a}
NaCl (2mM)+PS	42 ± 0.41^{b}	0.47 ± 0.21^{b}	4.2 ± 0.19^{b}
NaCl (4mM)+PS	31 ± 0.29^{b}	0.35 ± 0.14^{b}	3.1 ± 0.12^{b}
NaCl (6mM)+PS	$23\pm0.18^{\rm c}$	0.26 ± 0.07^{b}	2.3 ± 0.07^{c}

 Table 1. Effect of salt stress on seed germination, relative germination rate and germination index of Coriandrum sativumL. var. Kasturi with or without application of potassium silicate.

Values are mean \pm SE of three replicates from three independent experiments.

Table 2. Effect of salt stress on seedling length, biomass and vigour index of *Coriandrum sativum* L. var. Kasturi with or without application of potassium silicate.

Treatment	Radicle length (cm)	Plumule length (cm)	Fresh weight (mg/g)	Dry weight (mg/g)	Relative water content (%)	Vigour index
Control	5.62 ± 0.21^{a}	13.51 ± 0.41^a	4.22 ± 0.23^a	1.20 ± 0.08^a	$91.23{\pm}0.82^a$	16643.1 ^a
NaCl (6mM)	2.21 ± 0.17^{c}	5.10 ± 0.24^{c}	$1.14\pm0.18^{\rm c}$	0.29 ± 0.01^{c}	65.14 ± 0.21^{b}	877.2 ^d
PS (2mM)	$7.91{\pm}0.25^a$	14.22 ± 0.33^a	4.71 ± 0.52^a	1.59 ± 0.12^a	94.52 ± 0.37^a	20359.6 ^a
NaCl (2mM)+PS	6.12 ± 0.33^{a}	$11.24{\pm}0.62^{b}$	4.02 ± 0.23^a	0.98 ± 0.07^{b}	78.34 ± 0.24^{b}	7464.8 ^b
NaCl (4mM)+PS	5.54 ± 0.47^a	9.81 ± 0.72^{b}	3.76 ± 0.63^{b}	$0.91\ \pm 0.08^b$	$67.23{\pm}0.21^{\text{b}}$	5372.5 ^b
NaCl (6mM)+PS	$3.82{\pm}0.19^{b}$	7.56 ± 0.19^{c}	2.19 ± 0.44^{b}	0.72 ± 0.02^{b}	$58.12\pm0.15^{\rm c}$	2617.4 ^c

Values are mean \pm SE of three replicates from three independent experiments.

Chlorophyll content reflected significant increase with potassium silicate treatmentas compared to control and NaCl treatment. Total chlorophyll content reflected the following trend: PS > C > NaCl (2mM) + PS > NaCl (4mM) + PS > NaCl (6mM) + PS > NaCl. The carotenoid content was highest in NaCl (6mM) + PS treatment. It might be due to the protective role of carotenoid in coriander seeds against salt stress (Table 3). Similarly, Silicon improves the rigidity and erectness of leaves to enhance photosynthesis in sugarcane (Verma*et al.*2019). Reduction in photosynthesis in*Medicago truncatula*under salt stress could be due to the reduction in chlorophyll contents, damage of photosynthetic apparatus and inhibition of ribulose-1,5-bisphophate enzyme which resulted reduction in PSII efficiency (Najar*et al.*2019).

The cell membranes are first target of environmental stresses, due to this membrane damage was evaluated through electrolyte leakage. The electrolyte leakage was maximum (37%) in salt treated coriander seeds (Fig. 1). Malondialdehyde (MDA) is product of lipid peroxidation which showed the following trend: NaCl > NaCl (6mM) + PS >NaCl (4 mM) + PS > NaCl (2mM) + PS > C > PS (Fig. 1). The gradual increase in MDA content was observed in coriander seedlings with increase in NaCl concentration. Results revealed that potassium silicate treatment reduced MDA content by 59% signifying its protective role under salt stress. This result was in agreement with the work of Soleimannejad *et al.* (2019) who reported that silicon improved plasma membrane

activity by lowering electrolyte leakage via increasing H^+ -ATPase activity which helps in Na⁺ exclusion from plant tissues.

Table 3. Effect of salt stress on pigment content of Coriandrum sativum L. var. Kasturi with or without

Chlorophyll b Total Chlorophyll Treatment Chlorophyll a Carotenoids $(mg.g^{-1}FW)$ $(mg.g^{-1} FW)$ $(mg.g^{-1} FW)$ $(mg.g^{-1} FW)$ Control 1.82 ± 0.43^{a} 0.84 ± 0.07^{a} 2.66 ± 0.98^{a} 0.58 ± 0.03^{b} NaCl (6 mM) $0.87 \pm 0.37^{\circ}$ 0.31 ± 0.01^c $1.18{\pm}\,0.71^{c}$ 0.25 ± 0.01^{c} PS (2 mM) 0.93 ± 0.04^{a} 2.89 ± 0.52^{a} 0.62 ± 0.08^a 1.96 ± 0.58^{a} 1.18 ± 0.31^{b} 0.62 ± 0.02^b 1.80 ± 0.34^{b} 0.53 ± 0.06^{b} NaCl (2 mM) + PS 0.99 ± 0.62^{b} 0.45 ± 0.02^{b} 1.44 ± 0.21^{b} 0.64 ± 0.04^a NaCl (4 mM) + PS 0.93 ± 0.58^{b} $0.39 \pm 0.04^{\circ}$ NaCl (6 mM) + PS $1.32 \pm 0.14^{\circ}$ 0.71 ± 0.11^a



Fig. 1. Effect of salt stress on electrolyte leakage and lipid peroxidation of *Coriandrum sativum* L. var. Kasturi with or without application of potassium silicate.

Biochemical constituents such as sugar, proline and protein contents were also affected by salt stress. Maximum amount of sugar and protein contents were present in coriander leaves with potassium silicate treatment (Table 4). Similarly, compatible osmolytes stabilize enzymes and protein complexes under salt stress (Rajasheker *et al.* 2019). Proline content showed following trend: NaCl (6mM) + PS > NaCl (4mM) + PS > NaCl (2mM) + PS > NaCl > PS > C.

The impact of salt stress on total antioxidant content in the leaves of *Coriandrum sativum* L. var. Kasturiwas studied with or without application of potassium silicate. It showed following trend: NaCl (6mM) + PS > NaCl (4mM) + PS > NaCl (2mM) + PS > PS > NaCl > C. Maximum total antioxidant content 46% was recorded in NaCl (6mM) + PS treatment (Fig. 2). This result is supported by Yin *et al.* (2019) who found that silicon application enhanced polyamine

application of potassium silicate.

accumulation in cucumber plants which might play a significant function in modulation of antioxidant defense system by reducing oxidative stress, thus increasing the salt tolerance ability.

Table 4. Effect of salt stress on sugar, proline and protein contents of Coriandrum sativum L. var.

Kasturi with or without application of potassium silicate.				
Treatment	Sugar	Proline	Protein	
	$(mg.g^{-1}DW)$	$(\mathbf{u}\mathbf{M}.\mathbf{g}^{-1}\mathbf{D}\mathbf{W})$	$(mg.g^{-1}FW)$	

	$(mg.g^{-1}DW)$	$(\mu M.g^{-1} DW)$	$(mg.g^{-1}FW)$
Control	$2.54{\pm}0.72^{\text{b}}$	$16.2\pm0.12^{\rm c}$	10.42 ± 0.12^{b}
NaCl (6 mM)	2.05 ± 0.21^{c}	27.11 ± 0.56^{b}	$8.15\pm0.07^{\text{c}}$
PS (2 mM)	3.87 ± 0.54^a	26.32 ± 0.18^b	14.76 ± 0.37^a
NaCl (2 mM) + PS	2.83 ± 0.41^{b}	37.24 ± 0.42^a	10.15 ± 0.53^{b}
NaCl (4 mM) + PS	3.10 ± 0.39^a	4431 ± 0.35^a	9.52 ± 0.45^{b}
NaCl (6 mM) + PS	3.29 ± 0.22^{a}	47.49 ± 0.49^{a}	8.24 ± 0.09^{c}

Values are mean \pm SE of three replicates from three independent experiments.



Fig. 2. Effect of salt stress on total antioxidant content of *Coriandrum sativum* L. var. Kasturi with or without application of potassium silicate.

Potassium silicate utilization in crop fields to combat salinity is eco-friendly, cost-effective and sustainable approach. It contains both potassium and highly soluble silicon which is required for plant growth and its application does not release any hazardous by-products in nature. Hence, the use of potassium silicate can be recommended for the growth of coriander plants under saline conditions.

Salt stress significantly reduced seed germination and growth parameters of coriander. It exhibited electrolyte leakage and lipid peroxidation and reduced pigment, sugar and protein contents but enhanced proline accumulation. Potassium silicate supplementation mitigated unfavorable effects of salt stress in coriander by triggering the up-regulation of proline and total antioxidant activity.

More focus is required on the effects of potassium silicate under field conditions with multiple stresses rather than laboratory studies. Further in-depth studies are needed to explore biochemical nature of potassium silicate and potassium silicate mediated salt tolerance mechanism at the transcriptome, proteome and metabolome level. Potassium silicate utilization in crop fields to combat salinity is eco-friendly, cost-effective and sustainable approach. It contains both potassium and highly soluble silicon which are required for plant growth and its application does not release any hazardous by-products in nature. Hence, the use of potassium silicate can be recommended for the growth of coriander plants under saline conditions.

Acknowledgements

The authors are thankful to Amity Institute of Biotechnology, Amity University, Noida, India for offering laboratory facilities to carry out this study.

References

- Abdul Baki AA and Anderson JD 1973. Viability and leaching of sugars from germinating seeds by textile, leather and distillery industries. Indian J. Environ. Prot. **11**: 592 -594.
- Almeida D M, Oliveira MM and Saibo N J 2017. Regulation of Na⁺ and K⁺ homeostasis in plants: towards improved salt stress tolerance in crop plants. Genet. Mol. Biol. **40:** 326-345.
- Barrs HD and Weatherley PE 1962. A re-examination of the relative turgidity technique for estimating water deficits in leaves. Aust. J. Biol. Sci. **15**: 413-428.
- Bates LS, Waldren RP and Teare ID 1973. Rapid determination of free proline for water-stress studies. Plant Soil. **39**: 205-207.
- Chahal KK, Singh R, Kumar A and Bhardwaj U 2017. Chemical composition and biological activity of *Coriandrum sativum* L. a review. Indian J. Nat. Prod. Resour. **8**(3): 193-203.
- Farhangi-Abriz S and Torabian S 2018. Nano-silicon alters antioxidant activities of soybean seedlings under salt toxicity. Protoplasma. 255(3): 953-962.
- Garg N and Bhandari P 2016. Interactive effects of silicon and arbuscular mycorrhiza in modulating ascorbate-glutathione cycle and antioxidant scavenging capacity in differentially salt-tolerant *Cicer arietinum* L. genotypes subjected to long-term salinity. Protoplasma **253**: 1325-1345.
- Abdel Latef AAH, Akter A and Tahjib-Ul-Arif M 2021. Foliar application of auxin or cytokinin can confer salinity stress tolerance in *Vicia faba* L. Agronomy. 11(4):790.
- Guerriero G, Hausman JF and Legay S 2016. Silicon and the plant extracellular matrix. Front. Plant Sci. 7: 463-470.
- Hafez EM, El Dein Omara A, Alhumaydhi FA and El-Esawi MA 2021. Minimizing hazard impacts of soil salinity and water stress on wheat plants by soil application of vermicompost and biochar. Physiol. Plant. 172(2): 587-602.
- Hedge JE and Hofreiter BT 1962. Estimation of carbohydrate. In Whistler, R.L. and Be Miller, J.N. (Ed.), Methods in carbohydrate chemistry. Academic Press, New York. pp. 17-22.
- Hurtado AC, Chiconato DA, Prado RDM, Junior GDSS, Viciedo DO, Piccolo MDC 2020. Silicon application induces changes C : N : P stoichiometry and enhances stoichiometric homeostasis of sorghum and sunflower plants under salt stress. Saudi J. Biol. Sci.27: 3711-3719.
- Kirk JTO and Allen RL 1965. Dependence of chloroplast pigment synthesis on protein system: effect of actidione. Biochem.Biophys. Res. Commun. 21: 523-530.
- Laribi B, Kouki K, M'Hamdi M and Bettaieb T 2015.Coriander (*Coriandrum sativum* L.) and its bioactive constituents. Fitoterapia. **103**: 9-26.
- Li Y 2008. Effect of salt stress on seed germination and seedling growth of three salinity plants. Pak. J. Biol. Sci. 11: 1268-1272.

- Liu P, Yin L, Wang S, Zhang M, Deng X, Zhang S and Tanaka K 2015. Enhanced root hydraulic conductance by aquaporin regulation accounts for silicon alleviated salt-induced osmotic stress in *Sorghum bicolor* L. Environ. Exp. Bot. 111: 42-51.
- Lowry OH, Rosebrough NJ, Farr AL and Randall RJ 1951.Protein measurement with the Folin phenol reagent.J. Biol. Chem. 193: 265-275.
- Misyura M, Colasantil J and Rothstein SJ 2013. Physiological and genetic analysis of *Arabidopsis thaliana* anthocyanin biosynthesis mutants under chronic adverse environmental condition. J. Expt. Bot. **64**(1): 229-40.
- Najar R, Aydi S, Sassi-Aydi S, Zarai A and Abdelly C 2019. Effect of salt stress on photosynthesis and chlorophyll fluorescence in *Medicago truncatula*. Plant Biosys.-An Int. J. Dealing All Aspects Plant Biol. 153: 88-97.
- Prieto P, Pineda M and Aguilar M 1999. Spectrophotometric quantitation of antioxidant capacity through the formation of phosphor molybdenum complex: specific application to determination of vitamin E. Anal. Biochem. 269: 337-341.
- Rajasheker G, Jawahar G, Jalaja N, Kumar SA, Kumari PH, Punita D L and Kishor P BK 2019. Role and regulation of osmolytes and ABA interaction in salt and drought stress tolerance. In: Plant Signaling Molecules. Woodhead Publishing. pp. 417-436.
- Riaz M, Arif MS, Ashraf M A, Mahmood R, Yasmeen T and Shakoor M B 2019. A comprehensive review on rice responses and tolerance to salt stress. In:Advances in Rice Research for Abiotic Stress Tolerance. Elsevier. pp. 133-158.
- Saddiq MS, Iqbal S, Hafeez MB, Ibrahim AMH, Raza A, Fatima EM, Baloch H, Jahanzaib Woodrow P and Ciarmiello LF 2021.Effect of salinity stress on physiological changes in winter and spring wheat. Agronomy.11: 1193-1209.
- Safdar H, Amin A, Shafiq Y, Ali A, Yasin R, Shoukat A,Hussan M Ul and Sarwar MI 2019. A review: impact of salinity on plant growth. Nat. Sci. 17: 34-40.
- Shi Y, Zhang Y, Yao H, Wu J, Sun H and Gong H 2014. Silicon improves seed germination and alleviates oxidative stress of bud seedlings in tomato under water deficit stress. Plant Physiol.Biochem. 78: 27-36.
- Soleimannejad Z, Abdolzadeh A and Sadeghipour HR 2019. Beneficial effects of silicon application in alleviating salinity stress in halophytic *Puccinellia distans* plants. Silicon **11**: 1001-1010.
- Verma KK, Liu XH, Wu KC, Singh RK, Song QQ, Malviya MK, Song XP, Singh P, Verma CL and Li YR 2020.The impact of silicon on photosynthetic and biochemical responses of sugarcane under different soil moisture levels. Silicon. 1-13.12:1355-1367
- Wu J, Guo J, Hu Y and Gong H 2015. Distinct physiological responses of tomato and cucumber plants in silicon-mediated alleviation of cadmium stress. Front. Plant Sci. 6: 453-466.
- Yin J, Jia J, Lian Z, Hu Y,Guo J, Huo H, Zhu Y and Gong H 2019. Silicon enhances the salt tolerance of cucumber through increasing polyamine accumulation and decreasing oxidative damage. Ecotoxicol. Environ. Saf. 169: 8-17.
- Zargar SM, Mahajan R, Bhat JA, Nazir M and Deshmukh R 2019. Role of silicon in plant stress tolerance: Opportunities to achieve a sustainable cropping system. 3 Biotech. 9: 73.
- Zhu YX, Yin JL, Liang Y,Liu JQ, Jia JH, Huo HQ, Wu ZF, Yang RL and Gong HJ 2019. Transcriptomic dynamics provides an insight into the mechanism for silicon-mediated alleviation of salt stress in cucumber plants. Ecotoxicol. Environ. Saf. 174: 245-254.

(Manuscript received on 04 August 2021, revised on 09 August 2022)